Cytometer Cleaning

ICBR Cytometry

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Abstract

A well-cleaned instrument goes a long way toward insuring the reliability and long-term operability of our instruments. We appretiate your adherence to this protocol when starting up and shutting down. We love our users. Please contact us with any questions or concerns. Thank you, and please enjoy your cytometry.

1 Introduction

When cleaning the instrument, we address two modules of the fluidics. One module is addressed when the arm is out, and the other is addressed when the arm is in. Because of the high rate of flow when the arm is out, the steps in the protocol addressing the arm-out module are briefer in duration than the steps with the arm in. For all steps, you will be running with the flow rate of the cytometer on high, a setting that is modified by selecting the appropriate flow rate on the control panel located directly on the Fortessa or LSR II. If you have any questions about the protocol, please do not hesitate to reach out to our Twitter handle, ICBRCytometry, by email, ICBR-Cytometry@ad.ufl.edu, or by phone, (352) 273-8186.

2 Protocol

2.1 IMPORTANT Info

The SIT is the nozel that collects the sample for measurement. At no point should you allow the tube to run dry while the arm is under the tube. At no point should you allow the arm to be under an empty SIT.

2.2 Steps

1. Set the instrument to “HI” and “RUN”
2. Put a tube containing 5% contrad (or the contrad solution provided for you on the bench) on the SIT and allow it to run for 1 minute with the arm out.
3. Pivot the arm to beneath the tube and allow it to run for 1 minute.
4. Run the provided 30% bleach solution with the arm out for 1 minute.
5. Run the bleach solution with the arm in for 4 minutes.
6. Run deionized water with the arm out for 1 minute.
7. Run water with the arm in for 4 minutes.
8. Set the instrument to “LO” and “STNDBY”.